

Resi*Pure*™ *ADVANCED*

SOLUTION FOR PEPTIDE PURIFICATION







Optimize the Purification of your GLP-1 Agonists and Peptide Analogs

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ResiPure[™] ADVANCED High Resistance Silica for Optimized Peptide Purification

Resi*Pure Advanced* is a high-performance functionalized spherical silica specifically designed to meet the rigorous demands of peptide purification.

Engineered for both analytical and preparative chromatography, Resi*Pure* ADV (*ADVANCED*) offers unparalleled robustness, efficiency, reproducibility, and scalability, making it the ideal solution for applications in pharmaceutical, biotechnology, and research settings.

Resi*Pure* ADV is built to perform under challenging conditions. Its wide pH range tolerance for alkaline wash (pH 1–13) and compatibility with organic solvents, including trifluoroacetic acid (TFA), enable flexibility across diverse purification workflows. Moreover, its exceptional chemical and mechanical stability ensures durability over multiple cycles, reducing costs and downtime while maintaining high-quality results.



ResiPure ADVANCED Silica Bead

At the core of ResiPure ADV's performance is its optimized particle design.

The perfectly spherical silica particles ensure uniform packing, minimal back-pressure, and consistent flow, resulting in sharp and symmetrical peaks even for complex peptide mixtures. Its dense functionalization and advanced endcapping provide robust hydrophobic interactions, allowing for efficient separation of hydrophobic peptides with superior resolution.

Perfectly Spherical

High Purity & Low Metal Content

Dense Functionalization

High Mechanical & Chemical Resistance

Uniform Pore Size & Density



Why Choose ResiPure ADVANCED

Optimal Separation Performance

· Excellent selectivity and efficiency

pH 1 to 13 for alkaline wash

Material Stability & Robustness

- · High mechanical strength
- · High chemical resistance, with a wide pH stability

■ Batch-to-Batch Reproducibility

• Material and performance consistency, batch after batch

Combines all benefits of other gels on the market

Excellent ROI

- · High performance purification
- · Particle offering long lifetime

■ Compliance with Regulatory Requirements

- · Meets all pharmaceutical standards
- Full traceability and documentation: technical data sheets and certificates of analysis (CoA)

Renowned technical support

Manufactured by a Silica Specialist

- 30 years expertise in chromatography, purification and material science
- · Product designed, developped and supported by our own R&D team
- · Renowned technical and customer support

Easily Scalable

· Industrial capacities: from 10 g to multi-ton quantities

Metric tons available











Ordering Information

Bulk spherical silica gels

ResiPure ADV silica gels are offered from 10 g up to multi-ton scale.



ResiPure ADV C18				
SKU	Quantity			
S03107H-B-100G	100 g			
S03107H-B-250G	250 g			
S03107H-B-1KG	1 kg			
S03107H-B-5KG	5 kg			
S03107H-B-10KG	10 kg			
S03107H-B-25KG	25 kg			

ResiPure ADV C8					
SKU	Quantity				
S30907H-B-100G	100 g				
S30907H-B-250G	250 g				
S30907H-B-1KG	1 kg				
S30907H-B-5KG	5 kg				
S30907H-B-10KG	10 kg				
S30907H-B-25KG	25 kg				

ResiPure ADV C4				
SKU	Quantity			
S32807H-B-100G	100 g			
S32807H-B-250G	250 g			
S32807H-B-1KG	1 kg			
S32807H-B-5KG	5 kg			
S32807H-B-10KG	10 kg			
S32807H-B-25KG	25 kg			

HPLC columns

For analytical needs, HPLC columns packed with ResiPure ADV are also offered: contact us!



Available sizes:

- length: 50 mm, 100 mm, 150 mm, and 250 mm.
- diameter: 4.6 mm, 10 mm, 21.2 mm, 30 mm, and 50 mm.

Phases Portfolio

To meet all purification needs, Resi*Pure* ADV is offered in three functionalizations:

		C18	C8	C4
PN		S03107H-B	S30907H-B	S32807H-B
Structure		(s) ~~~~~~~~	3 ~~~~	3 ~~
Material		Silica	Silica	Silica
Particle Size		10 μm	10 μm	10 μm
Pore Diameter		150 Å	150 Å	150 Å
Specific Surfa	ce Area	250 m²/g	250 m²/g	250 m²/g
Endcapping		Yes	Yes	Yes
nU Stability	Mobile Phase	1 - 10	1 - 10	1 - 10
pH Stability	Washing	up to pH 13	up to pH 13	up to pH 13
Description		Most hydrophobic, best for small to medium-sized peptides, provides high retention and resolution.	Moderate hydrophobicity, suitable for larger or moderately hydrophobic peptides, offering shorter retention times than C18.	Least hydrophobic, ideal for large or highly hydrophobic peptides and proteins, reducing retention and peak broadening. Generaly requires more aqueous eluents.



Batch-to-Batch Reproducibility

The spherical shape and uniform particle size distribution minimize back-pressure and ensure smooth column packing, reducing the risk of channeling or uneven flow. This uniformity translates to consistent performance, maintaining separation efficiency and peak resolution over multiple cycles.

Hydrophobic retention profiles on 3 different batches of ResiPure ADV CHROMATOGRAPHIC TEST CONDITIONS Detection: UV at 254 nm Column: C18, 250 \times 4.6 mm, 10 μm Mobile phase: acetonitrile/water (80/20) Injection volume: 5 µL Sample: acetophenone (1), toluene (2), naphtalene (3), and anthracene (4) Flow rate: 1.000 mL/min Column temperature: 25°C ResiPure ADV C18 16000 Batch #1 14000 ResiPure ADV C18 Batch #2 10000 ResiPure ADV C18 Batch #3 1- Acetophenone 2- Toluene 3- Naphtalene 4- Anthracene ■ ResiPure ADV C18 ■ Resi*Pure* ADV C18 ■ ResiPure ADV C18 Batch #3 Efficiency and retention were similar for the hydrophobic markers analysed on 3 different batches of ResiPure ADV.

Morphology

Scanning Electron Microscope (SEM) particles pictures of 3 batches of ResiPure ADV.

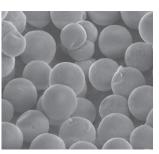
Batch #1



Batch #2



Batch #3





Peptide Analysis on ResiPure ADV C18

Sigma-Aldrich H2016 Peptide Mix

CONDITIONS

Column: C18, 250 × 4.6 mm, 10 μm

Mobile phases

- Mobile phase A (MPA): 0.1 % TFA in water (v/v)
- Mobile phase B (MPB): 0.1 % TFA in acetonitrile (v/v)

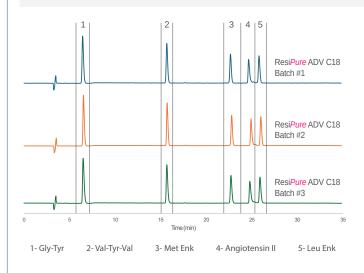
Gradient

- 1. Hold 3 minutes at 90/10 (MPA/MPB)
- 2. Increase to 65/35 (MPA/MPB) during 27 minutes
- 3. Hold 5 minutes at 65/35 (MPA/MPB)

Flow rate: 1.000 mL/min Column temperature: 25°C Detection: UV at 220 nm Injection volume: 10 μL

Sample : calibration peptide mixture (Sigma Aldrich H2016) diluted in 1 mL of

water.





From one batch to the other, retention, selectivity and efficiency were found to be the same for the different compounds in a peptide mixture.

Brunauer-Emmett-Teller (BET) Analysis

X.		BET Analysis			
Batch	Surface area (m²/g)	Pore size (A)	Pore volume (mL/g)	Carbon Load (%)	
1	250	147	0.92	17.5	
2	239	154	0.92	16.5	
3	248	149	0.93	17.0	

BET analysis shows uniformity in material and chemical properties of ResiPure ADV silica particles from batch to batch.

Resi*Pure* ADV offers reproducibility and consistency of its particles, batch after batch.



Superior Mechanical Resistance

By choosing our robust silica, you benefit from a reduced cost of operation, as fewer replacements and less downtime mean optimized productivity. The durability of our material ensures that you get the most out of every batch, making it a cost-effective solution.

Test: Packing - Repacking of a Preparative Column

CONDITIONS

Test Method

Procedure: 20 consecutive cycles of column packing / unpacking

Column: C18, 150×21.2 mm, $10 \mu m$ Mobile phase: acetonitrile/water (70/30)

Flow rate: 15 mL/min

Detection: UV at 254 nm

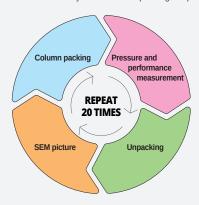
System: Shimadzu Preparative HPLC

Injection volume: $500~\mu\text{L}$

Sample: mixture of acetophenone, toluene, naphthalene, and anthracene (marker)

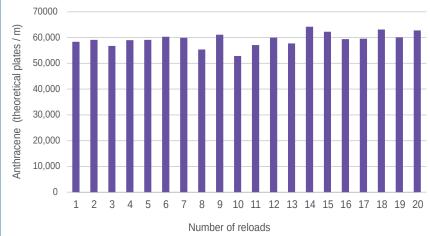
Test Procedure

20 consecutive cycles of column packing / unpacking



Results

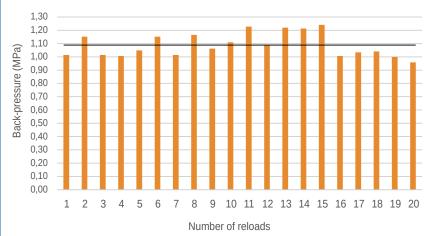
Column performance vs number of reloads





Results

Back pressure vs number of reloads



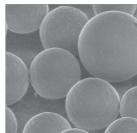
SEM Pictures

Resi*Pure* ADV particles SEM pictures at the start, and after several repacking processes:

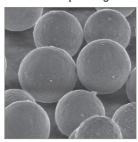
Before



After 10 packings



After 20 packings



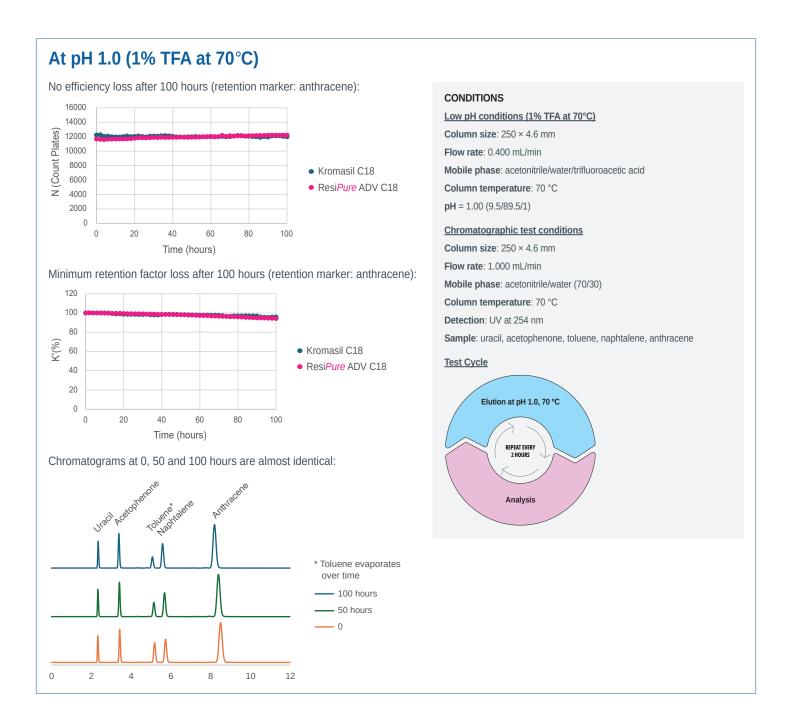
The silica beads are still perfectly spherical, with smooth surfaces free of any cracks or cavities, even after packing and unpacking 20 times a column of 21.2 mm ID.



pH Stability in Mobile Phase

pH stability in the mobile phase is essential for consistent and reproducible peptide separation. Fluctuations in pH can lead to changes in peptide charge, affecting their interaction with the stationary phase and thus the separation process.

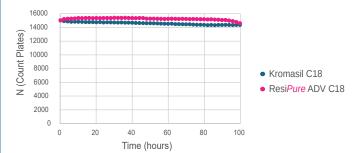
A stable pH ensures that peptides maintain their desired charge state throughout the chromatographic process, optimizing separation efficiency.



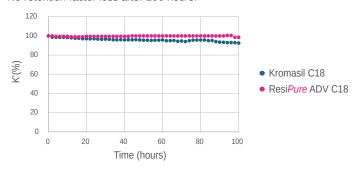


At pH 10.0 (Ammonium carbonate at RT)

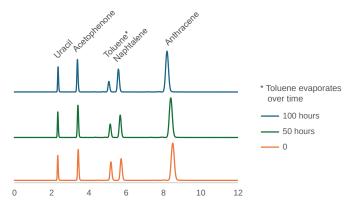
No efficiency loss after 100 hours:



No retention factor loss after 100 hours:



Chromatograms at 0, 50, and 100 hours are perfectly identical:



CONDITIONS

pH 10 conditions

Column size: 250 × 4.6 mm Flow rate: 0.400 mL/min

Mobile phase: 10 mM ammonium carbonate in 90/10 (acetonitrile/water),

adjusted to pH 10.00

Column temperature: 23 °C

Chromatographic test conditions

Column size: 250 × 4.6 mm Flow rate: 1.000 mL/min

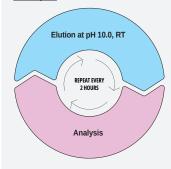
Mobile phase: acetonitrile/water (80/20)

Column temperature: 23 °C

Detection: UV at 254 nm

Sample: uracil, acetophenone, toluene, naphtalene, anthracene

Test Cycle



Resi*Pure* ADV can be run from pH 1 to 10 without compromising your results.



pH Stability through Washing Cycles

pH stability in the mobile phase is essential for consistent and reproducible peptide separation. Fluctuations in pH can lead to changes in peptide charge, affecting their interaction with the stationary phase and thus the separation process.

A stable pH ensures that peptides maintain their desired charge state throughout the chromatographic process, optimizing separation efficiency.

pH 1 to 13 for alkaline wash

Test: Alkaline wash cycles (pH 13)

CONDITIONS

Alkaline washing cycle conditions

Column: C18, 250 × 4.6 mm, 10 μm

Flow rate: 1.000 mL/min

Cycles

- Step 1 Alkaline wash (pH 13): ACN/0.1 M NaOH (50/50), 6 CV
- Step 2 Neutralisation (pH 3.5): ACN/water adjusted to pH 3.5 with glacial acetic acid (50/50), 6 CV
- Step 3 Wash: 100% acetonitrile, 6 CV
- Step 4 Column performance analysis (marker: anthracene)

Alkaline wash REPEAT UNTIL DETERIORATION Column performance analysis Wash

Chromatographic test conditions

Column: C18, 250 × 4.6 mm, 10 μm **Mobile phase**: acetonitrile/water (80/20

Mobile phase: acetonitrile/water (80/20) Flow rate: 1.000 mL/min

Column temperature: 25°C

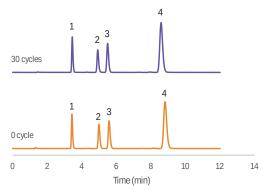
Detection: UV at 254 nm

Test compound: anthracene

Sample: acetophenone (1), toluene (2), naphtalene (3), and anthracene (4)

Results

Performance after 30 cycles

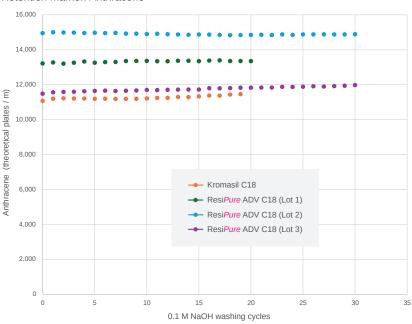




Results

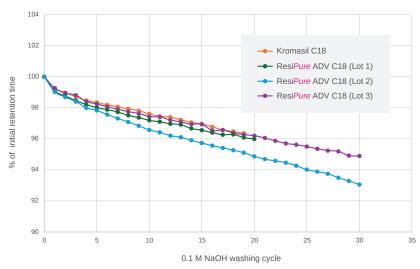
Loss of efficiency after alkalin washing cycle (pH 13)

Retention marker: Anthracene



Loss of retention after alkalin washing cycle (pH 13)

Retention marker: Anthracene



ResiPure ADV for washing cycles:

- · alkaline resistance, alkaline durability, alkaline-tolerant
- wide pH tolerance range, up to pH 13
- enduring more than 30 cycles of NaOH washing without showing any meaningful back-pressure increase / performance loss



ResiPure ADV C18 vs Kromasil® C18 Analysis of Thymalfasin from a synthetic crude product by C18 liquid chromatography

Known for its immunomodulatory properties, Thymalfasin is a synthetic 28-amino acid peptide derived from thymosin alpha-1. It has a linear structure with an N-terminal acetylation, which enhances its stability.

With a molecular weight of approximately 3108.3 g/mol, it is a hydrophilic peptide, highly soluble in water due to its multiple acidic residues, such as aspartic and glutamic acids.

CONDITIONS

Column: 250 × 4.6 mm, 10 μm

Mobile phases

• Mobile phase A (MPA): 50 mM ammonium formate in water, buffered at pH 3.8

· Mobile phase B (MPB): Acetonitrile

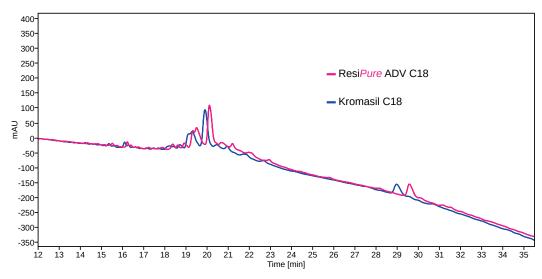
Flow rate: 1.000 mL/min Column temperature: 25°C Detection: UV at 215 nm

Sample load: 40 μL of 50 mg crude product diluted in 10 mL of 1% of acetic acid in

water (v/v)

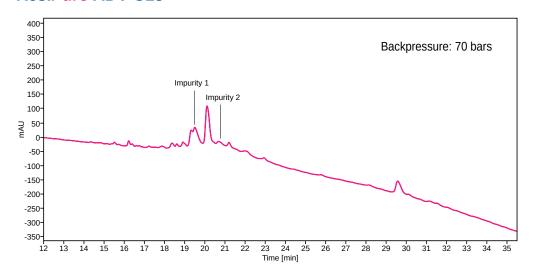
4	Gradient				
Time (min)	MPA (%)	MPB (%)			
0.01	95	5			
5.00	95	5			
65.00	20	80			
70.00	20	80			
70.01	95	5			
80.00	95	5			

Impurities profiles comparison: ResiPure ADV C18 vs Kromasil® C18

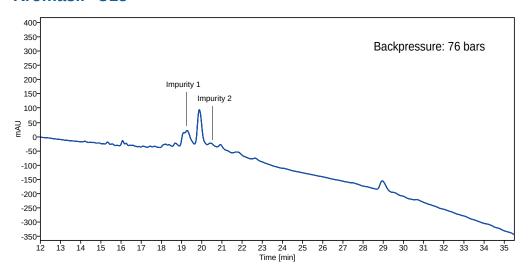




ResiPure ADV C18



Kromasil® C18



Comparison of the two chromatograms						
Stationary phase	Retention time (min) Plate Selectivity Selectivity Selectivity 1) (Thymalfasin-Impurity 1) (Thymalfasin-Impurity 2) (Thymalfasin-Impurity 1) (Thymalfasin-Impurity 2)					
Kromasil® C18	19.87	45 133	1.04	1.03	1.27	1.57
Resi <i>Pure</i> ADV C18	20.10	55 355	1.04	1.03	1.30	1.63

Resi*Pure* ADV is more efficient than the Kromasil® C18 to separate Thymalfasin from the main impurities in a crude product, while generating less back pressure.



ResiPure ADV C18 vs Kromasil® C18 Analysis of Tirzepatide from a synthetic crude product by C18 liquid chromatography

Tirzepatide is a synthetic peptide consisting of 39 amino acids, designed as a dual GIP and GLP-1 receptor agonist. It has a linear structure with a fatty acid moiety, enhancing plasma stability and albumin binding.

CONDITIONS

Columns: 250×4.6 mm, $10 \mu m$

Mobile phases

· Mobile phase A (MPA): 10 mM ammonium acetate in water

· Mobile phase B (MPB): acetonitrile

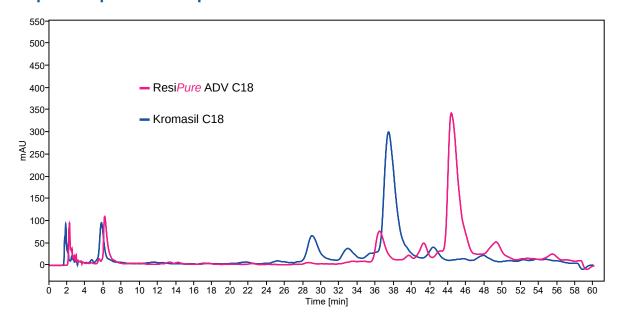
Flow rate: 1.000 mL/min Column temperature: 25°C Detection: UV at 220 nm

Sample load: 5 µL of 120 mg of crude product diluted in 5 mL of dissolving solution

Dissolving solution: 50/50 acetonitrile/water

Gradient					
Time (min)	MPA (%)	MPB (%)			
0.01	60	40			
50.00	53	47			
55.00	53	47			
55.01	60	40			
60.00	60	40			
80.00	95	5			

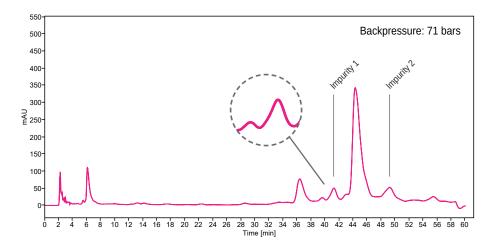
Impurities profiles comparison: ResiPure ADV C18 vs Kromasil® C18



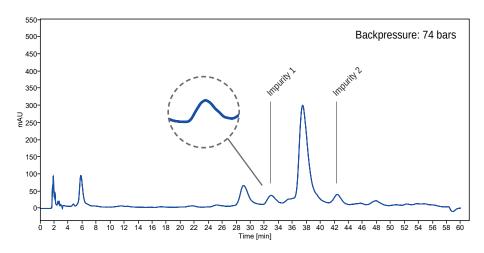


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ResiPure ADV C18



Kromasil® C18



Comparison of the two chromatograms						
Stationary phase	Retention time (min) Plate (Tirzepatide) Selectivity Selectivity Selectivity (Tirzepatide-Impurity 1) (Tirzepatide-Impurity 2) (Tirzepatide-Impurity 1) (Tirzepatide-Impurity 1)					
Kromasil® C18	37.44	4 715	1.15	1,14	2.16	2,29
Resi <i>Pure</i> ADV 18	43.10	8 486	1.08	1.12	1.81	2.38

A slightly different selectivity is observed when using ResiPure ADV C18 compared to Kromasil® C18.

Resi*Pure* ADV C18 allows a better selectivity with Tirzepatide than Kromasil® C18 for some impurities.

On the other hand, Kromasil® C18 allows a better selectivity for other impurities.



Retention profile for a peptide mixtures

A calibration peptides mixture, Sigma-Aldrich H2016, composed of Angiotensin II, Gly-Tyr, Leu enkephalin, Met enkephalin and Val-Tyr-Val, has been tested with Resi*Pure* ADV C18 to evaluate its retention efficiency.

CONDITIONS

Column: C18, 250 × 4.6 mm, 10 µm

Mobile phases

- Mobile phase A (MPA): 0.1 % TFA in water (v/v)
- Mobile phase B (MPB): 0.1 % TFA in acetonitrile (v/v)

Gradient

- 1. Hold 3 minutes at 90/10 (MPA/MPB)
- 2. Increase to 65/35 (MPA/MPB) during 27 minutes
- 3. Hold 5 minutes at 65/35 (MPA/MPB)

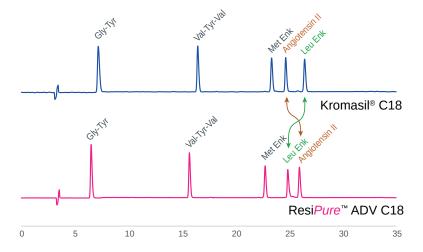
Flow rate: 1.000 mL/min

Column temperature: 25°C

Detection: UV at 220 nm **Injection volume**: 10 μ L

Sample: calibration peptide mixture (Sigma Aldrich H2016) diluted in 1 mL of water.

Chromatogram





Efficiency for each peptide component



Resi*Pure* ADV can offer different selectivity compared to Kromasil®, while providing high efficiency.



pH Effect

The pH of a solution plays a crucial role in peptide separation. Adjusting pH helps optimize the interaction between peptides and the stationary phase, improving purification outcomes.

pH variation effect on Tirzepatide

CONDITIONS

Column: C18, 250 × 4.6 mm, 10 µm

Mobile phases

- · Mobile phase A (MPA):
 - Test 1: 50 mM ammoniun formate in water, adjusted to pH 3.5
 - o Test 2: 10 mM potassium phosphate dibasic in water, adjusted to pH 8.0
- · Mobile phase B (MPB): acetonitrile

Gradient

- 1. Hold 10 minutes at 68/32 (MPA/MPB)
- 2. Increase to 55/45 (MPA/MPB) during 50 minutes
- 3. Hold 10 minutes at 55/45 (MPA/MPB)
- 4. Reequilibrate at 68/32 (MPA/MPB) for 10 minutes

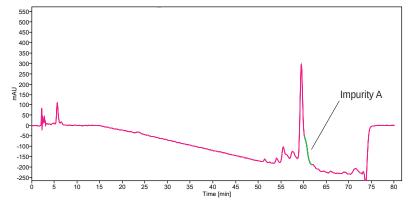
Flow: 1.000 mL/min

Column temperature: 25°C Detection: UV at 220 nm

Injection volume: 5 µL

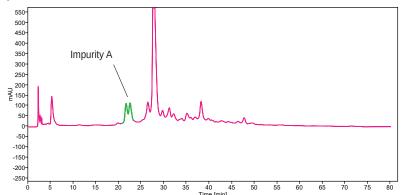
Sample: 125 mg of crude material diluted in 5 mL of acetonitrile/water (50/50)

pH 3.5



Tirzepatide elutes at 59.4 minutes, with some impurities clustered before and after the main peak.

pH 8.0



Tirzepatide elutes at 27.6 minutes, with a greater number of well-resolved impurities appearing afterwards.



Comparison with competitor at pH 8.0 Kromasil® vs ResiPure ADV 500-450-400 350 300 250 — Kromasil® 200 150· 100· 50 -50 -100· -150 -200 -250 40 45 Time [min] 550 500 450· 400 350 300 250 - ResiPure ADV ¥ 150 100 50 -50--150 -200 -250· 40 45 Time [min] 450 400 ResiPure ADV 350 300 250 — Kromasil® 200-150-100 50 -50 -100· -150· -200 -250-ResiPure ADV provides better separation of Comparison of the two chromatograms Tirzepatide from its impurities compared to Kromasil®. Stationary phase Retention time (min) Kromasil® C18 23.914 4113 1.08 0.78 ResiPure ADV C18 27.612 8301 1.08 1.61

A pH change in eluting conditions on reverse chromatography can lead to a favorable selectivity, and thus obtain the desired separation between impurity and targeted peptide.



Resi*Pure* ADV C18 vs Resi*Pure* ADV C8 Change of stationary phase selectivity

For peptide purification applications, it is strongly recommended to verify the selectivity of both C8 and C18. Depending on the hydrophobicity of the peptide, using a C8 instead of a C18 can improve selectivity under the same elution conditions.

CONDITIONS

Columns: C18 & C8, 250 \times 4.6 mm, 10 μm

Mobile phases

- Mobile phase A (MPA): 10 mM potassium phosphate dibasic in water, adjusted to pH 8.0
- · Mobile phase B (MPB): acetonitrile

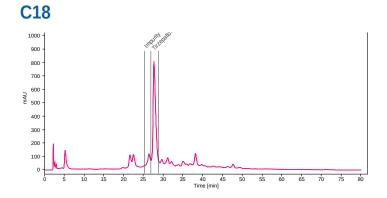
Gradient

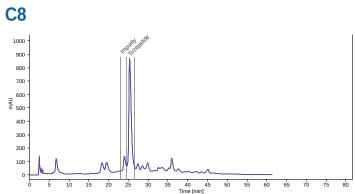
- 1. Hold 10 minutes at 68/32 (MPA/MPB)
- 2. Increase to 55/45 (MPA/MPB) during 50 minutes
- 3. Hold 10 minutes at 55/45 (MPA/MPB)
- 4. Reequilibrate at 68/32 (MPA/MPB) for 10 minutes

Flow rate: 1.000 mL/min Column temperature: 25°C Detection: UV at 220 nm Injection volume: 5 µL

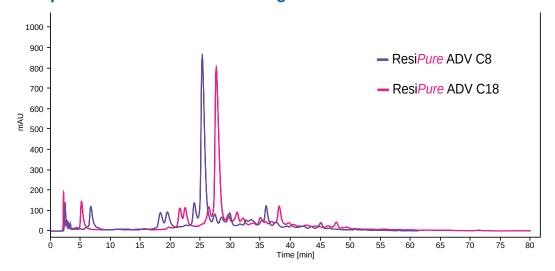
Sample: 125 mg of crude material diluted in 5 mL of acetonitrile/water (50/50)

4	Gradient				
Time (min)	MPA (%)	MPB (%)			
0.01	68	32			
10.00	68	32			
60.00	55	45			
70.00	55	45			
70.01	68	32			
80.00	68	32			





Comparison of the two chromatograms



C8 vs C18 for Tirzepatide Analysis					
Concretion nevernators	Resi <i>Pure</i> Stat	ionnary Phase			
Separation parameters	C8	C18			
α _{Tirzepatide-Impurity}	1.06	1.05			
R _{Tirzepatide-Impurity}	1.27	1.01			

This application shows that ResiPure[™] ADV C8 stationary phase is more selective than ResiPure[™] ADV C18 for purifying Tirzepatide from crude material, with a lower retention time under the same eluent conditions.



Gradient Effect

The gradient effect optimizes separation by gradually modifying the mobile phase composition. This enhances selectivity, improves peak resolution, and minimizes impurities.

By fine-tuning gradient conditions, process chemists achieve high-purity peptides efficiently, ensuring superior results.

Here, we compare two gradients, of the same duration but varying ACN percentage.

Comparison of two gradients

CONDITIONS

Column: ResiPure ADV C8, 250 × 4.6 mm, 10 µm

Mobile phases

- Mobile phase A (MPA): 10 mM buffer phosphate pH 8.00
- Mobile phase B (MPB): 100% acetonitrile

Flow: 1.000 mL/min

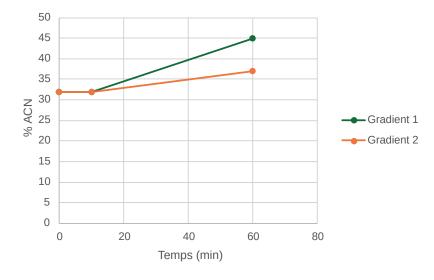
Column temperature: 25°C

Detection: UV at 215 nm

Injection volume: $5~\mu L$

Sample: 125 mg of crude material diluted in 5 mL of acetonitrile/water (50/50)

Varying ACN percentage



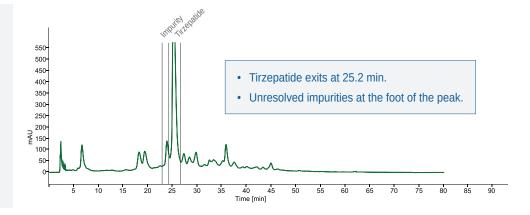


Gradient 1

• 0 min: 32% ACN

• 10 min: 32% ACN

• 60 min: 45% ACN

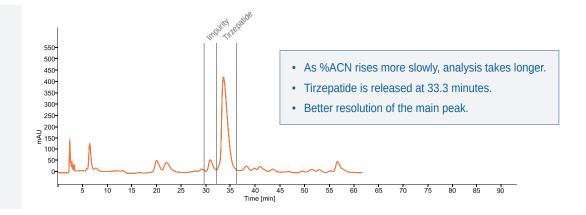


Gradient 2

• 0 min: 32% ACN

• 10 min: 32% ACN

• 60 min: 37% ACN



Gradient Effect			
Separation parameters	Gradient		
	1	2	
α _{Tirzepatide-Impurity}	1.06	1.09	
R _{Tirzepatide-Impurity}	1.27	1.49	

Change of linear gradient conditions can lead to an improvement in selectivity to separate an API from the other impurities.



Purification of Tirzepatide

The crude material has a purity about 22.3% in Tirzepatide. A purification process must be developed which requires a multi-step purification to achieve the final target purity of >99%.

Analytical purity test

The purity of the different extract was determined according to the following HPLC conditions.

LIQUID CHROMATOGRAPHY CONDITIONS

Column: SiliaChrom dT C18, 250 × 4.6 mm, 3 μm

Flow: 1.000 mL/min

Column Temperature: 30 °C

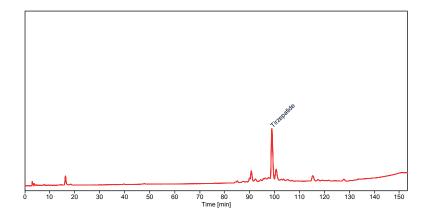
Gradient

• MPA: 0.1% of trifluoroacetic acid in water (v/v)

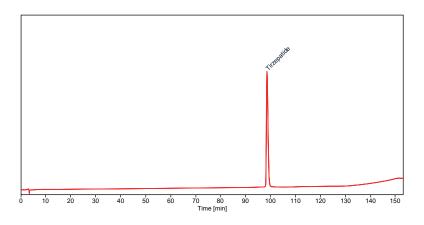
• MPB: 0.1% of trifluoroacetic acid in Acetonitrile (v/v)

Detection: UV at 220 nm

Gradient		
MPA (%)	MPB (%)	
60	40	
53	47	
53	47	
60	40	
60	40	
95	5	
	MPA (%) 60 53 53 60 60	



Liquid chromatographic profile of Tirzepatide for the crude material: 20 mg/mL diluted in 75/25 (Water/Acetonitrile), injection Volume: 3 μ L.



Liquid chromatographic profile of Tirzepatide for the reference standard: 1 mg/mL diluted in 50/50 (Water/Acetonitrile), injection Volume: 20 $\mu L.$

Purification process summary

Two successive purification steps were necessary to purify the Tirzepatide from the crude material to get the targeted purity (>99%).

- Both purification steps were performed in using the Resi*Pure* ADV C8.
- The first purification step was performed under the basic conditions (pH 8.00).
- The second purification step was performed under the acidic conditions (pH 3.50)

Refer to figure 2 and 3.

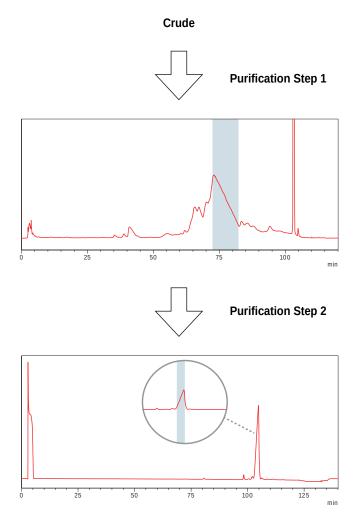


Figure 2: Purification process summary of the Tirzepatide

Initial purity: 22.3% 110 120 **Purification Step 1** Purity: 91% Yield: 95% 110 120 **Purification Step 2** Purity: 99.8% Yield: 72% 100 110 120

Figure 3: Tirzepatide purity analysis by liquid chromatography for each purification step.



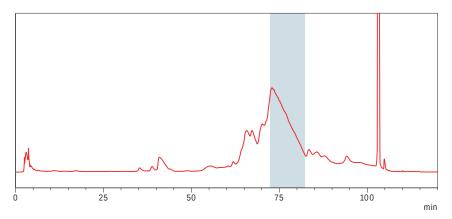
Purification step 1: removal of bulk impurities

The initial purification step was carried out with the crude Tirzepatide product. The purity of the crude material was about 22.3%.

The method development was performed at analytical scale screening the complete Resi*Pure* ADV stationary phases. The best results regarding separation efficiency, target purity and recovery rate were obtained with Resi*Pure* ADV C8 in using the elution conditions fixed to pH 8.00.

The final fractionation was optimised to get a target purity of 91% and a yield of 95% starting from a very low purity initially 22.3% of the crude. This chromatographic purification step at pH 8.00 with the Resi*Pure* ADV leads to a massive improvement of the purity level of Tirzepatide.

The conditions of the purification step 1 are described below:



CONDITIONS

Column: Resi*Pure* ADV C8, 250 × 10 mm, 10 µm

Eluent

A) 20 mM NH₄HCO₃, pH 8.00

B) Acetonitrile

Gradient: 25%B (0-10 min), 25-35%B (10-80 min), 35%B (80-100 min), 80%B (100-110 min), and 25%B (110-120 min)

Flow rate: 4.75 mL/min Column temperature: 25°C Detection: UV at 220 nm Injection volume: 2mL

Sample: 40 mg/mL Tirzepatide crude in 75/25 (water/acetonitrile)

Collection: 72.7 minutes to 82.0 minutes



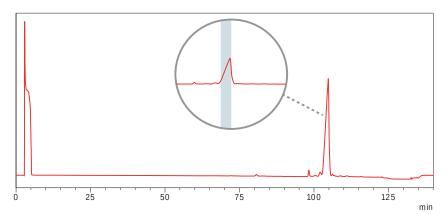
Purification step 2: final polishing

The second chromatography step in this purification process was a polishing step.

This step was also carried out with the ResiPure ADV C8. The purification was performed at pH 3.5 leading to great separation results.

This second step leads to a final product purity of 99.8% with a yield about 75%.

The conditions of the purification step 2 are described below:



CONDITIONS

Column: Resi*Pure* ADV C8, 250 \times 10 mm, 10 μ m

Eluent

A) 50 mM NH₄HCO₃, pH 3.5

B) Acetonitrile

Gradient: 20%B (0-10 min), 20-30%B (10-60 min), 30-55%B (60-120 min), 80%B (120-130 min) and 20%B (130-140 min)

Flow rate: 4.75 mL/min Column temperature: 25°C Detection: UV at 220 nm

Injection volume: 1 mL (25 mg load)

Sample: Tirzepatide crude after the purification step 1, 25 mg/mL in DMSO

Collection: 103.5 minutes to 105.00 minutes



Conclusion

Two successive purification steps were necessary to purify the Tirzepatide from the crude material to the targeted purity (>99.5%).

- Both purification steps were performed in using the Resi*Pure* ADV C8.
- The first purification step was performed under the basic conditions (pH 8.00) to remove bulk impurities from the crude.
- The second purification step was performed under the acidic conditions (pH 3.50) to polish the extract coming from the first purification step.

In using this process, the purity of the Tirzepatide in the final extract was 99.8%. The purity of Tirzepatide in the starting material was 22.3 %.

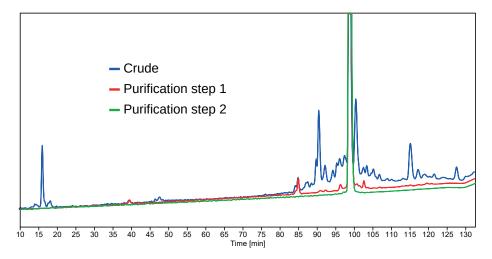


Figure 4: Overlay of the different chromatographic profiles for Tirzepatide analyzed after each purification step.



About SiliCycle



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